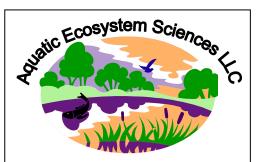
#### TECHNICAL MEMORANDUM

# Tenmile Lakes Toxic Algae Monitoring, September 21, 2009

Prepared for: Tenmile Lakes Basin Partnership

Prepared by: Jacob Kann, Ph.D.

Date: September 29, 2009



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September 21<sup>st</sup>, 2009 toxic algal cell count results for Tenmile Lakes are as follows (see Figure 1 below for sample station location):

Station	Date	Microcystis (cells/ml)	Gloeotrichia (cells/ml)	Total Microcystis+ Gloeotrichia	Total <i>Anabaena</i> (cells/ml)
S3	9/21/2009	12,362	0	12,362**	260
S8	9/21/2009	1,901	0	1,901*	184
Z	9/21/2009	1,008,139	0	1,008,139***	17,932
N11	9/21/2009	4,126	0	4,126**	428
N16	9/21/2009	3,479	0	3,479**	1,603

Exceeds World Health Organization Alert Level 1 increased vigilance guideline level of 500 cells ml<sup>-1</sup> for potentially toxigenic species in drinking water systems.

Samples form September 21<sup>st</sup> show that M*icrocystis aeruginosa* (MSAE) at the standard stations (S3, S8, N11, and N16) generally continued to increase, with all stations except S8 (which declined) exceeding the WHO Alert Level 2 guideline of 2000 cells/ml (at which time DHS and local health services typically issue a public alert for drinking water lakes and reservoirs) (Table 1; Figure 2). In past years such levels have been associated with microcystin toxin levels exceeding the WHO 1 µg/L drinking water level in Tenmile Lakes, thus, home-owners should ensure that treatment systems are operating effectively. This would be especially true for S3 with a value of 12,363 cells/ml of MSAE. Total *Anabaena* continued to decline overall, and was below Alert Level 2 at all stations (Figure 2).

However, as noted in the two previous algal memos, during the course of sampling at the standard stations, it became apparent that patchy concentrations of blue-green algae were occurring in other areas of the lake (see Sep 8 and Sep 15 bloom memos). During the Sep 21<sup>st</sup> sample date an additional sample was collected from one of these more concentrated bloom areas located in Coleman Arm and is shown as sample station "Z" above in Table 1 and in the map in Figure 1. Results from this station clearly show a very high concentration of MSAE, with a cell density of >1 million cells/ml (Table 1). Such MSAE levels can be associated with microcystin toxin levels that constitute a high probability of adverse health effects for recreational users of the lake. State of Oregon posting guidelines for recreational water bodies are 40,000 cells/ml for *Microcystis* and 100,000 cells/ml for *Anabaena*.

Blue-green algae (cyanobacteria) continued to comprise the majority of algal biomass at S3, S8, and N11, and N16; with the predominant species being *Microcystis*, Aphanizomenon flos-aquae and Anabaena planktonica (Appendix I).

Exceeds World Health Organization Alert Level 2 public health posting guideline level of 2000 cells ml<sup>-1</sup> for potentially toxigenic species in drinking water systems

<sup>\*\*\*</sup>Exceeds State of Oregon Recreational Guideline Levels of 40,000 cells/ml for *Microcystis* or 100,000 cell/ml for *Anabaena* 

These results show that blue-green blooms are prevalent in varying areas of both North and South Tenmile Lakes, and that given both cell density results from this date and microcystin toxin results from Sep 8<sup>th</sup> that exceeded guidelines for recreation posting, I would continue to advise against utilizing lake water for potable or recreational purposes at his time.

Due to the patchy nature of blue-green algal blooms it is possible for higher Microcystis aeruginosa and Anabaena flos-aquae densities (and therefore higher microcystin toxin and anatoxin concentrations to be present in areas not sampled in this survey, particularly along shorelines or during calm conditions of little to no wind. Given the lakes' demonstrated history of toxic blooms, and the fact that all areas of the lake cannot be tested at all times, those utilizing the lake for drinking water should <u>always</u> follow Oregon Health Division recommendations for purification (attached). In addition, recreational users should <u>always</u> avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance. Moreover, because pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.

#### **References for Alert Levels**

- Carey, C.C., and J.F. Haney, and K.L. Cottingham. 2007. First report of microcystin-LR in the cyanobacterium *Gloeotrichia echinulata*. Environmental toxicology 22:337-339.
- Kann, J. 2007. Tenmile Lakes Toxic Algal Sampling Program: 2006 Data Summary Report. Tenmile Lakes Basin Partnership, Lakeside OR 97520
- Falconer et al. 1999. Safe levels and safe practices. Pages 155-177 *in*: I. Chorus and J. Bartram, editors. *Toxic Cyanobacteria in water: a guide to their public health consequences.* World Health Organization Report. E & FN Spon, London and New York.
- Stone, D., and W. Bress. 2007. Addressing Public Health Risks for Cyanobacteria in Recreational Freshwaters: The Oregon and Vermont Framework. Integrated Environmental Assessment and Management; 3(1): 137-143 (2007). http://www.oregon.gov/DHS/ph/hab/docs/Stone\_cyano\_rec.pdf
- Yoo, S.R., W.W. Carmichael, R.C. Hoehn, and S.E. Hrudy. 1995. Cyanobacterial (blue-green algal) toxins: a resource guide. AWWA Research Foundation and American Water Works Association. Denver, CO. 229 p. (ISBN 0-89867-824-2)

### Year 2009 Tenmile Lakes Sample Site Locations

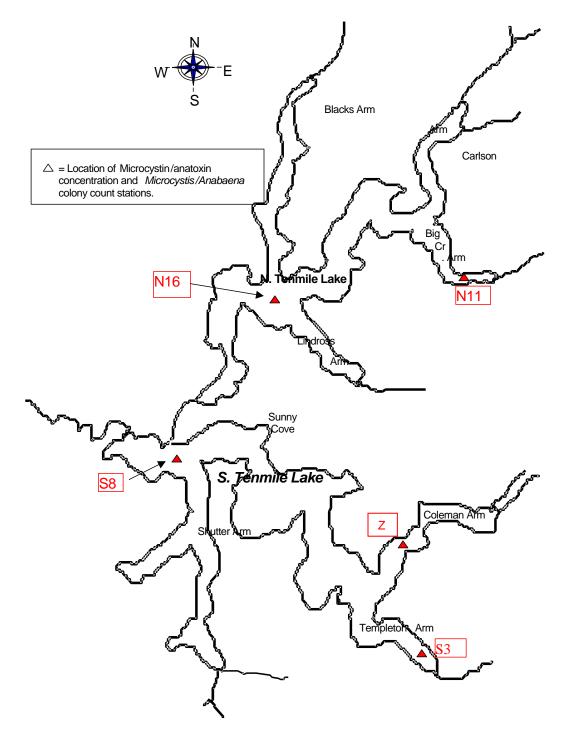


Figure 1. Sample station location for potentially toxigenic cyanobacteria, Tenmile Lakes, 2009.

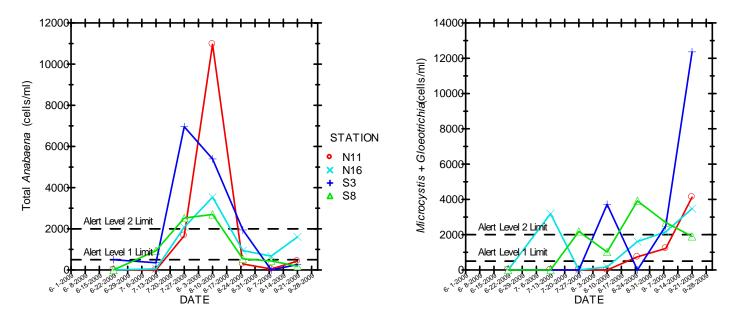


Figure 2. Trend of potentially toxic cyanobacteria in Tenmile Lakes, 2009.

# Appendix I: Aquatic Analysts Phytoplankton Lab Sheets

	Phytoplankto	on Sample Ana	alysis			
		Tenmile Lake				
	Sample Site:	S8				
	Sample Depth:					
	Sample Date:	21-Sep-09				
	Total Density (#/mL):	27				
	Total Biovolume (um³/mL):	51,924				
	Trophic State Index:	28.6				
		Density	Density	Biovolume	Biovolume	
	Species	#/mL	Percent	um³/mL	Percent	Group
	Microcystis aeruginosa	7.9	29.3	· · · · · · · · · · · · · · · · · · ·		bluegreen
	Anabaena planctonica	6.5	24.3			bluegreen
	Ochromonas sp.	3.1	11.6		0.5	chrysophyte
	Rhodomonas minuta	2.5	9.4	51	0.1	cryptophyte
	Asterionella formosa	1.0	3.9		0.4	diatom
	Dinobryon sertularia	1.0	3.9	125	0.2	dinoflagellate
7	Sphaerocystis schroeteri	0.7	2.8	104	0.2	green
8	Ankistrodesmus falcatus	0.7	2.8	19	0.0	green
9	Cocconeis placentula	0.7	2.8	342	0.7	diatom
10	Aphanizomenon flos-aquae	0.6	2.2	375	0.7	bluegreen
11	Cryptomonas erosa	0.4	1.7	232	0.4	cryptophyte
12	Melosira ambigua	0.4	1.7	1,052	2.0	diatom
13	Fragilaria crotonensis	0.3	1.1	750	1.4	diatom
14	Chlamydomonas sp.	0.1	0.6	48	0.1	green
15	Fragilaria capucina mesolepta	0.1	0.6	114	0.2	diatom
16	Glenodinium sp.	0.1	0.6	104	0.2	dinoflagellate
	Melosira italica	0.1	0.6	280	0.5	diatom
18	Trachelomonas volvocina	0.1	0.6	280	0.5	euglenoid
	Note: 200 count.					
	Microcystis aeruginosa cells/mL =	1,901				
	e.eeyee asraginosa sons/iiie =	1,001				
	Anabaena planctonica cells/mL =	177				
	Anabaena planctonica heterocysts/mL =	7				
	Anabaena planctonica akinetes/mL =	0.4				
	Aphanizomenon flos-aquae cells/mL =	6				
	Aquatic Analysts			Sample ID:	MB71	

	Phytoplankto					
		Tananila Laba				
		Tenmile Lake				
	Sample Site:	53				
_	Sample Depth:	04.0 00				
	Sample Date:	21-Sep-09				
	Total Density (#/mL):	76				
	Total Biovolume (um³/mL):	207,383				
	Trophic State Index:	38.5				
		Donoity	Density	Pievelume	Biovolume	
	0			um³/mL		0
	Species	#/mL	Percent	um <sup>9</sup> /mL	Percent	Group
	Microsystic compiles of	24.7	AF F	00.000	47.7	hlu a nua a n
	Microcystis aeruginosa	34.7	45.5	98,893		bluegreen
	Apphanizomenon flos-aquae	13.5 9.8	17.6 12.8	,		bluegreen
	Anabaena planctonica Ochromonas sp.			46,649 556		bluegreen
	Dinobryon sertularia	6.5 4.1	5.3	539		chrysophyte dinoflagellate
	Fragilaria crotonensis	2.9	3.7	38,433		diatom
	Sphaerocystis schroeteri	2.9		1,601	0.8	
	Ankistrodesmus falcatus	1.0	1.6	31	0.0	green
	Asterionella formosa	0.4		719	0.0	green diatom
	Synedra ulna	0.4	0.5	813	0.3	diatom
	Staurastrum gracile	0.4	0.5			
			0.5	221	0.1	green diatom
	Fragilaria vaucheriae Rhodomonas minuta	0.4	0.5	235 8	0.1	
13	KIIOGOIIIOIIAS IIIIIGIA	0.4	0.5	0	0.0	cryptophyte
	Note: 200 count.					
	Microcystis aeruginosa cells/mL =	12,362				
	Aphanizomenon flos-aquae cells/mL =	297				
٩pl	nanizomenon flos-aquae heterocysts/mL =	6				
	Anabaena planctonica cells/mL =	255				
	Anabaena planctonica celis/mL =  Anabaena planctonica heterocysts/mL =					
	Anabaena planctonica neterocysts/mL =  Anabaena planctonica akinetes/mL =	0.4				
	Aliabaella pianctonica akinetes/IIIL =	0.4				
_						
	Aquatic Analysts			Sample ID:	MB72	

_	Phytoplankto					
	. пусорияния					
	Sample:	Tenmile Lake				
	Sample Site:	N11				
	Sample Depth:					
	Sample Date:	21-Sep-09				
	Total Density (#/mL):	127				
	Total Biovolume (um³/mL):	197,659				
	Trophic State Index:	38.2				
	·					
		Density	Density	Biovolume	Biovolume	
	Species	#/mL	Percent	um³/mL	Percent	Group
_						
1	Aphanizomenon flos-aquae	47	37.1	50,373	25.5	bluegreen
	Microcystis aeruginosa	25	19.7	33,007	16.7	bluegreen
	Anabaena planctonica	19	15.0	76,702	38.8	bluegreen
	Cocconeis placentula	10	7.5	4,382	2.2	diatom
	Chlamydomonas sp.	5	3.8	1,548	0.8	green
	Melosira granulata	4	3.3	11,232	5.7	diatom
	Melosira ambigua	4	2.8	7,364	3.7	diatom
8	Cryptomonas erosa	2	1.4	929	0.5	cryptophyte
	Eunotia pectinalis	1	0.9	857	0.4	diatom
	Fragilaria crotonensis	1	0.9	6,001	3.0	diatom
1	Melosira granulata angustissima	1	0.9	595	0.3	diatom
	Gomphonema angustatum	1	0.9	214	0.1	diatom
	Navicula cryptocephala	1	0.9	220	0.1	diatom
	Sphaerocystis schroeteri	1	0.5	333	0.2	green
	Rhodomonas minuta	1	0.5	12	0.0	cryptophyte
6	Oocystis pusilla	1	0.5	129	0.1	green
7	Gomphonema subclavatum	1	0.5	357	0.2	diatom
	Synedra ulna	1	0.5	1,185	0.6	diatom
	Gloeocystis ampla	1	0.5	152	0.1	green
	Achnanthes minutissima	1	0.5	30	0.0	diatom
1	Fragilaria capucina mesolepta	1	0.5	1,518	0.8	diatom
	Asterionella formosa	1	0.5	393	0.2	diatom
3	Cosmarium sp.	1	0.5	125	0.1	green
	Note: 200 count.					
	Aphanizomenon flos-aquae cells/mL =	800				
p	hanizomenon flos-aquae heterocysts/mL =	2				
	Anabaena planctonica cells/mL =	419				
	Anabaena planctonica heterocysts/mL =	8				
	Anabaena planctonica akinetes/mL =	1				
_	Microcystis aeruginosa cells/mL =	4,126				
_					MDTO	
	Aquatic Analysts			Sample ID:	MB/3	

Ph	ytoplankt	on Sample Ana	alysis			
		- 1 1 1				
		Tenmile Lake				
	mple Site:	N16				
	ple Depth:	04.0 00				
Sar	nple Date:	21-Sep-09				
Total Dens	sity (#/ml \	713				
Total Biovolume						
Trophic St		51.5				
Tropine St	ate muex.	31.3				
		Density	Density	Biovolume	Riovolume	
Species			Percent	um³/mL	Percent	Group
Species		#/111L	Percent	uiii /iiiL	reiceili	Group
1 Aphanizomenon flos-aquae		580	81.3	730,693	58.3	bluegreen
2 Anabaena planctonica		55		272,209		bluegreen
3 Microcystis aeruginosa		23		27,836		bluegreen
4 Fragilaria crotonensis		14		170,495		diatom
5 Melosira ambigua		12	1.6	25,276		diatom
6 Asterionella formosa		12	1.6	13,268	1.1	diatom
7 Oocystis lacustris		3		893	0.1	green
8 Chlamydomonas sp.		3		942	0.1	green
9 Anabaena circinalis		3		6,176	0.5	bluegreen
10 Cryptomonas erosa		3	0.4	1,508	0.1	cryptophyte
11 Ochromonas sp.		3		246	0.0	chrysophyte
12 Melosira granulata angustissim	na	3	0.4	3,624	0.3	diatom
Note: 200 count.						
Aphanizomenon flos-aquae		11,598				
Aphanizomenon flos-aquae hetero	cysts/mL =	9				
Anabaena planctonica		1,487				
Anabaena planctonica hetero		26				
Anabaena planctonica akii	netes/mL =	3				
Minnessette	II-/I	0.470				
Microcystis aeruginosa	cells/mL =	3,479				
Anabaena circinalis	colle/ml =	87				
Anabaena circinalis	CEIIS/IIIL =	07				
Aquatic Analysts				Sample ID:	MB74	
1						

	Phytoplankto	on Sample Ana				
		Tenmile Lake				
	Sample Site:	Z				
	Sample Depth:					
_	Sample Date:	21-Sep-09				
H	Total Density (#/mL):	9,237				
	Total Biovolume (um³/mL):	15,823,810				
	Trophic State Index:	69.8				
	Hopino dato maexi	33.3				
		Density	Density	Biovolume	Biovolume	
	Species	#/mL	Percent	um³/mL	Percent	Group
	Microcystis aeruginosa	4,166	45.1	8,065,112	51.0	bluegreen
2	Aphanizomenon flos-aquae	3,577	38.7	4,056,555	25.6	bluegreen
	Anabaena planctonica	589	6.4	3,231,714	20.4	bluegreen
	Chlamydomonas sp.	226	2.5	73,582	0.5	green
_	Asterionella formosa	136	1.5	38,851	0.2	diatom
6	Melosira ambigua	136	1.5	184,027	1.2	diatom
	Rhodomonas minuta	91	1.0	1,811	0.0	cryptophyte
	Ankistrodesmus falcatus	91	1.0	2,264	0.0	green
	Tabellaria fenestrata	45	0.5	108,675		diatom
	Nitzschia frustulum	45	0.5	5,434		diatom
_	Achnanthes linearis	45	0.5	5,977		diatom
	Pinnularia sp.	45	0.5	18,112	0.1	diatom
13	Glenodinium sp.	45	0.5	31,697	0.2	dinoflagellate
	Note: 200 count.					
	Microcystis aeruginosa cells/mL =	1,008,139				
	Microcystis aeruginosa celis/IIIL =	1,006,139				
	Aphanizomenon flos-aquae cells/mL =	64,390				
Αp	hanizomenon flos-aquae heterocysts/mL =	181				
	Anabaena planctonica cells/mL =	17,660				
-	Anabaena planctonica celis/mL =  Anabaena planctonica heterocysts/mL =	272				
<u> </u>	Anabaena pianetonica neterocysts/mb =	212				
	Aquatic Analysts			Sample ID:	MB75	

**Oregon Health Division Drinking water treatment guidance** August 31, 2001

**DHS Contact Information:** 

Harmful Algae Program Coordinator: Laura Boswell at (971) 673 – 0438 If she is not available call the main line for the Office of Environmental

Public Health at: (971) 673 – 0440 or

Toll Free: (877) 290 – 6767 and press 0. Ask for Laura Boswell

- 1. Treatment systems should consist of sand filtration followed by chlorination, followed by activated charcoal filtration. It is essential that sand filtration be done before disinfection to remove as many algal cells as possible without killing or rupturing them.
- 2. Chlorination systems should be capable of maintaining at least 1 ppm of chlorine residual for at least 20 minutes contact time before the water enters the activated charcoal system.
- 3. The final step in the process should be effective activated charcoal treatment to remove toxin remaining after the sand filtration and disinfection processes.
- 4. All treatment equipment used should meet NSF standard 53, and should be adequately sized to treat the maximum amount of water that you use. Treatment equipment needs regular monitoring and servicing to assure that it functions properly.
- 5. Ideally all water entering your home should be treated as recommended. It is possible to treat only water used in the kitchen, but this increases chances that animals or pets would inadvertently drink untreated water.

As more monitoring is done and toxin levels are measured this advisory may be altered. The advisory is to remain in effect until specifically changed or lifted by county and state health officials.

#### **FACT SHEET**

#### TOXIC MICROCYSTIS BLOOMS IN TENMILE LAKES

(information modified from Oregon Health Division Document: Hazards from *Microcystis aeruginosa* in Fresh Water – http://www.ohd.hr.state.or.us/esc/docs/mafact.htm)

# What is a toxic bloom of Microcystis aeruginosa?

Microcystis aeruginosa is a species of blue-green algae that grows naturally in many surface waters. In most bodies of fresh water and most weather conditions it does not pose a hazard to wildlife or human beings. However, under certain conditions (such as when the water is warm with abundant nutrients) Microcystis aeruginosa can grow more rapidly than normal. The result can be excessive numbers of large colonies that form floating masses on the water surface or that are dispersed within the water column. These occurrences are called "algal blooms". Microcystis aeruginosa can produce natural toxins (called microcystins) that are very potent, and these toxins are higher in concentration during bloom conditions. The microcystin toxins are produced and contained inside the Microcystis cells, and are released to the water when the cells die and disintegrate. Also, since the cells are very small, they can be ingested along with the water. Toxin levels in a water body tend to be higher near shorelines and at the surface of the water where animal and human contact is most likely.

# What are the primary toxic effects of these blooms?

The primary toxic effect of microcystins is on the liver. At very high doses, death of liver cells and destruction of blood vessels in the liver can result in serious injury and possibly death. Though less is known about the long-term effects of microcystin toxins, animal studies have shown these toxins can cause chronic liver damage and may promote the formation of liver tumors. These effects are more likely to occur if exposure is frequent over a long period of time.

The levels of toxin necessary to produce immediate or acute illness in humans and animals are much higher than levels that may cause chronic liver injury. Drinking water standards are usually based on chronic effects. Currently, there is no drinking water standard in the U.S. for microcystins. Canada, Australia, and Great Britain have developed a guideline level of 1 microgram toxin per liter of water, or 1 part per billion (1 ppb). During algal blooms, toxin levels can greatly exceed 1 ppb.

# How is it determined when the water becomes safe once a bloom is reported?

Changes in weather or in other conditions in a water body influence the growth of blue-green algae. Generally, cooler weather, rainfall, and reduced sunshine will lead to reductions in algal growth and toxin levels. Algal blooms generally peak and die off rapidly and toxin levels in the water decline over days or weeks. Only blue-green algae experts can distinguish visually between different kinds of algal growth, and are able to determine when blooms have disappeared. Testing of the water is the only way to be certain that toxin levels are no longer dangerous.

## > When does the Oregon Health Division Issue Warnings?

**Drinking Water --** When measured or estimated toxin levels reach 1 ug/l the Department of Human Services, Office of Public Health Systems issues public advisories or warnings. These will include warnings regarding the use of water for drinking or food preparation unless the water has been treated following specific guidelines for destroying and removing toxins. Animals should be kept away from water during periods when microcystin toxin levels exceed 1 ug/l, because drinking the water can cause serious or even fatal illness.

**Contact Recreation** -- If levels are high enough to pose hazards for swimming, water-skiing or other direct skin contact activities, the advisories will warn against water contact. Generally skin hazards occur where the water has a green or blue-green color or where there are visible clumps or mats of algae present in the water. When measured toxin levels reach 5 ug/L or cell counts reach 15,000 cells/ml, contact recreation is considered unsafe.

# Can testing ensure that all areas of the lake are safe?

No, due to the patchy nature of blue-green algal blooms it is possible for higher *Microcystis* densities (and therefore higher microcystin toxin concentrations) to be present in areas not sampled in a given survey, particularly along shorelines or during calm conditions of little to no wind. Therefore, when a lake has a demonstrated history of algal toxicity or the presence of known toxin producing algal species, those utilizing the lake for drinking water should always follow Oregon Health Division recommendations for purification. In addition, recreational users should always avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance.

### Are domestic animals at risk during blooms?

Yes, pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.

### Is it safe to eat fish and other aquatic life?

Clams, mussels, snails and other shellfish should not be eaten during microcystin advisory periods, but it is believed that fish can be safely eaten if they are cleaned and all internal organs discarded. Internal organs of such fish may be toxic even to animals.

## How much does testing cost?

Samples must be shipped to qualified laboratories for analysis. A microscopic determination to quantify the number of Microcystis colonies and cells costs \$90 per sample. A specialized test to analyze for the microcystin toxin concentration costs \$100 per sample (overnight shipping costs not included), and for anatoxins the cost is \$250/sample.

 NOTE: A fact sheet about microcystin toxin and its effects may be found on the Web at <u>www.dhs.state.or.us/publichealth/esc/docs/mafact.cfm</u>