

Jacob Kann, Ph.D.  
Aquatic Ecologist

295 East Main St., Suite 7  
Ashland, OR 97520  
Voice: 541-482-1575  
Fax: 541-552-1024  
Email: [jacobkann@aol.com](mailto:jacobkann@aol.com)



## Tech Memo

To:	Mike Mader/Jason – TLBP	From:	Jake Kann
Fax:	(503) 731-4077 541-759-3711	Pages:	12
Phone:		Date:	09-18-07
Re:	<i>Tenmile Sampling</i>	CC:	Ken Kauffman – OHD

To all concerned,

September 12<sup>th</sup>, 2007 toxic algal cell count results for Tenmile Lakes are shown in Table 1:

Table 1. Cell density of potentially toxigenic cyanobacteria in Tenmile Lakes.

Station <sup>1</sup>	Date	<i>Microcystis</i> (cells/ml)	<i>Gloeotrichia</i> (cells/ml)	Total <i>Microcystis</i> + <i>Gloeotrichia</i> <sup>2</sup>	Total <i>Anabaena</i> (cells/ml)
S3	9/12/2007	1,877	0	1,877	3,162
S8	9/12/2007	131	0	131	8,687
N11	9/12/2007	6,605	0	6,605	304
N16	9/12/2007	69	0	69	2,818

<sup>1</sup>See Figure 1 for station location

<sup>2</sup>The combined density of MSAE and GTEC is computed because GTEC is a potential microcystin producer (Carey et al. 2007).

Samples collected on September 12<sup>th</sup> show that cell density of potentially toxigenic cyanobacteria species increased at several stations for both *Microcystis aeruginosa* (MSAE) and total *Anabaena* (*A. flos-aquae* [ABFA]+ *A. planctonica* [ABPL]) (Table 1; Figure 2). MSAE density increased at both of the stations located in the arms (S3 and N11; Figure 1), with S3 exceeding the WHO Alert Level 1 guideline (500 cells ml<sup>-1</sup> or increased vigilance level for drinking water systems), and N11 at 6,605 cells/ml exceeding the WHO Alert Level 2 guideline of 2,000 cells ml<sup>-1</sup> (at which time DHS and local health services typically provide notification to remind homeowners to follow water treatment guidelines—see Appendix II). Past research from Tenmile Lakes as well as other lake systems indicates that 5000 cells per ml of MSAE can commonly be associated with 1 µg/L of the hepatotoxin microcystin.

Levels of total *Anabaena* remained similar to the previous sample period at both N11 and N16; with N16 continuing to exceed the Alert Level 2 guideline (Figure 2). However, increases in total *Anabaena* occurred at both south-Lake stations, with S8 sharply increasing to 8,687 cells per ml (exceeding the Alert Level 2 guideline by 4.3 times). However, as stated in previous Tech Memos, the predominant species was *Anabaena planctonica* (Appendix I), a species less commonly associated with toxin production.

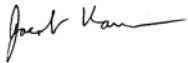
September 18, 2007

Biovolume at stations S3, S8, and N16 continued to be comprised mainly of the cyanobacterium *Anabaena planctonica*, which represented 59.4, 86.5, and 81.3 percent of the total biovolume (Appendix I). North-Lake station N11 was comprised approximately equally by MSAE (23.2%) and ABPL (23.7%). Another cyanobacterium (*Aphanizomenon flos-aquae*) not known to produce cyanotoxins in Oregon comprised 32.8% of the biovolume at S3 (Appendix I). Various Chrysophytes, Cryptophytes, and Diatoms comprised the remainder of the biovolume at all stations.

Given the continued presence and slightly increasing trend (red line; Figure 2) of potentially toxigenic cyanobacteria in Tenmile Lakes that exceeded Alert Level 2 (particularly the maximum MSAE value of 6,605 cells/ml at N11), and the fact that cyanobacterial cells have been reported in home-owner drinking water treatment systems (see Kann 2007), all drinking water protection efforts should be in place. However, levels of all potentially toxic cyanobacteria were well below Oregon's recreational guidelines (100,000 cells per ml for *Anabaena* and 40,000 cells/ml for *Microcystis*).

Please call if you have any questions.

Sincerely,



Jacob Kann Ph.D.  
Aquatic Ecologist

*Due to the patchy nature of blue-green algal blooms it is possible for higher Microcystis aeruginosa and Anabaena flos-aquae densities (and therefore higher microcystin toxin and anatoxin concentrations to be present in areas not sampled in this survey, particularly along shorelines or during calm conditions of little to no wind. Given the lakes' demonstrated history of toxic blooms, and the fact that all areas of the lake cannot be tested at all times, those utilizing the lake for drinking water should always follow Oregon Health Division recommendations for purification (attached). In addition, recreational users should always avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance. Moreover, because pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.*

---

#### References for Alert Levels

Carey, C.C., and J.F. Haney, and K.L. Cottingham. 2007. First report of microcystin-LR in the cyanobacterium *Gloeotrichia echinulata*. *Environmental toxicology* 22:337-339.

Kann, J. 2007. Tenmile Lakes Toxic Algal Sampling Program: 2006 Data Summary Report. Tenmile Lakes Basin Partnership, Lakeside OR 97520

Falconer et al. 1999. Safe levels and safe practices. Pages 155-177 in: I. Chorus and J. Bartram, editors. *Toxic Cyanobacteria in water: a guide to their public health consequences*. World Health Organization Report. E & FN Spon, London and New York.

Yoo, S.R., W.W. Carmichael, R.C. Hoehn, and S.E. Hrudy. 1995. Cyanobacterial (blue-green algal) toxins: a resource guide. AWWA Research Foundation and American Water Works Association. Denver, CO. 229 p. (ISBN 0-89867-824-2)

Year 2007 Tenmile Lakes Sample Site Locations

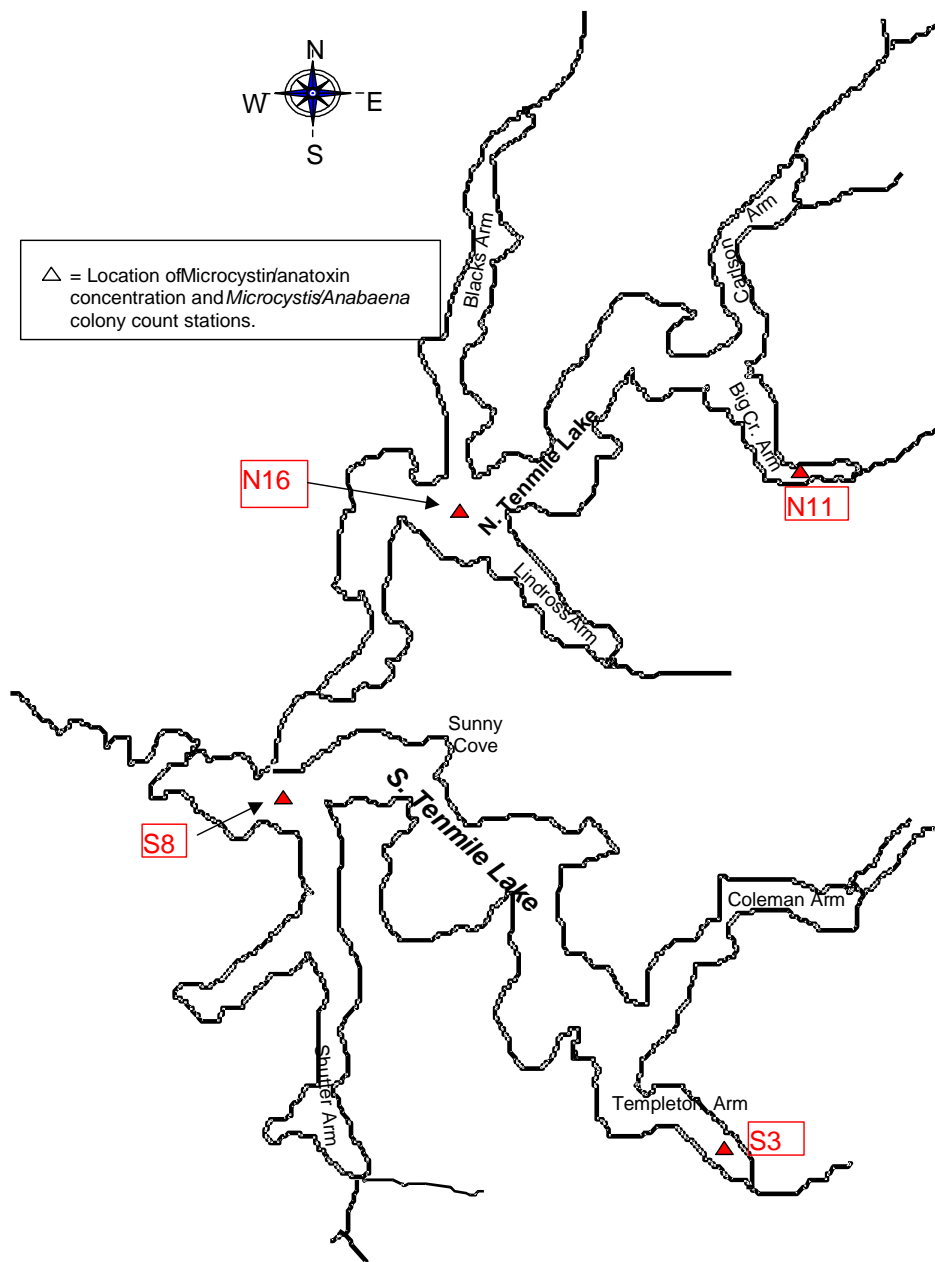


Figure 1. Sample station location for toxic cyanobacteria sampling in Tenmile Lakes, 2007.

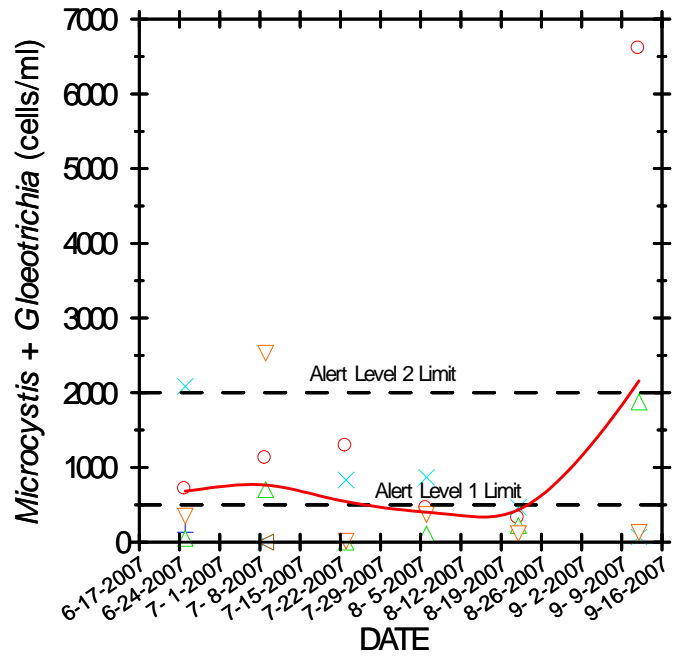
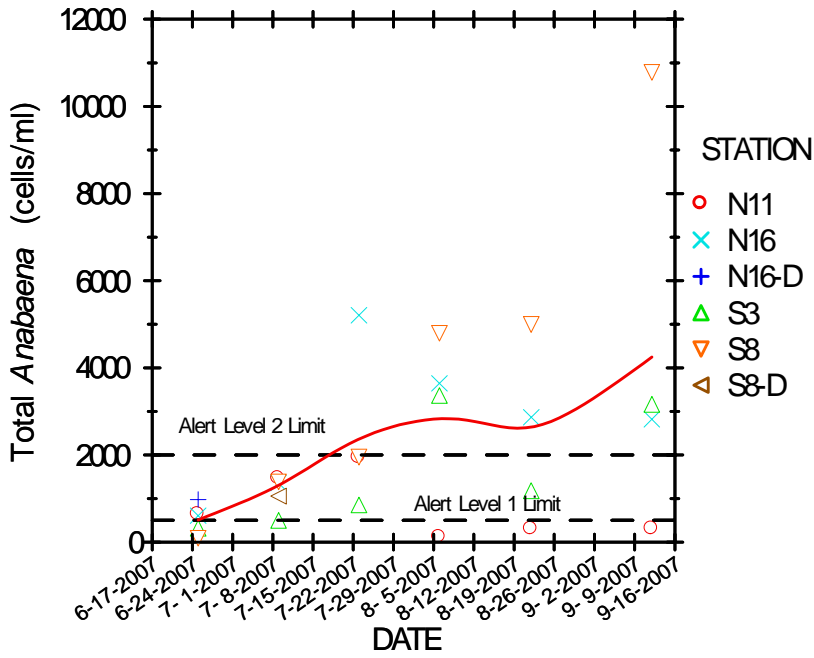


Figure 2. Trend of potentially toxic cyanobacteria in Tenmile Lakes (red line is distance weighted least square trend line), 2007.

## Appendix I – Aquatic Analysts Laboratory Reports

Phytoplankton Sample Analysis					
<b>Sample:</b>		Tenmile Lake			
<b>Sample Station:</b>		S3			
<b>Sample Depth:</b>					
<b>Sample Date:</b>		12-Sep-07			
<b>Total Density (#/mL):</b>		326			
<b>Total Biovolume (um<sup>3</sup>/mL):</b>		966,792			
<b>Trophic State Index:</b>		49.6			
<b>Species</b>	<b>Density #/mL</b>	<b>Density Percent</b>	<b>Biovolume um<sup>3</sup>/mL</b>	<b>Biovolume Percent</b>	<b>Group</b>
1 Aphanizomenon flos-aquae	194	59.5	317,117	32.8	bluegreen
2 Anabaena planctonica	101	31.1	574,108	59.4	bluegreen
3 Melosira granulata	15	4.5	39,527	4.1	diatom
4 Microcystis aeruginosa	6	1.8	15,019	1.6	bluegreen
5 Melosira ambigua	4	1.4	18,919	2.0	diatom
6 Cocconeis placentula	1	0.5	675	0.1	diatom
7 Chlamydomonas sp.	1	0.5	477	0.0	green
8 Cyclotella meneghiniana	1	0.5	557	0.1	diatom
9 Nitzschia dissipata	1	0.5	395	0.0	diatom
Anabaena planctonica cells/mL =		3,137			
Anabaena planctonica heterocysts/mL =		25			
Aphanizomenon flos-aquae cells/mL =		5,034			
Aphanizomenon flos-aquae heterocysts/mL =		97			
Microcystis aeruginosa cells/mL =		1,877			
<b>Aquatic Analysts</b>					
			<b>Sample ID:</b> KR74		



Phytoplankton Sample Analysis					
<b>Sample:</b> Tenmile Lake					
<b>Sample Station:</b> N11					
<b>Sample Depth:</b>					
<b>Sample Date:</b> 12-Sep-07					
<b>Total Density (#/mL):</b> 149					
<b>Total Biovolume (um<sup>3</sup>/mL):</b> 227,299					
<b>Trophic State Index:</b> 39.2					
<b>Species</b>	<b>Density #/mL</b>	<b>Density Percent</b>	<b>Biovolume um<sup>3</sup>/mL</b>	<b>Biovolume Percent</b>	<b>Group</b>
1 Achnanthes minutissima	29	19.8	1,769	0.8	diatom
2 Gomphonema angustatum	18	11.9	3,821	1.7	diatom
3 Cocconeis placentula	12	7.9	5,426	2.4	diatom
4 Fragilaria capucina mesolepta	12	7.9	19,550	8.6	diatom
5 Microcystis aeruginosa	10	6.9	52,840	23.2	bluegreen
6 Melosira ambigua	9	5.9	19,799	8.7	diatom
7 Eunotia pectinalis	7	4.5	5,255	2.3	diatom
8 Anabaena planctonica	6	4.0	53,961	23.7	bluegreen
9 Gomphonema subclavatum	6	4.0	6,015	2.6	diatom
10 Cryptomonas erosa	4	3.0	2,300	1.0	cryptophyte
11 Scenedesmus quadricauda	4	3.0	1,150	0.5	green
12 Chlamydomonas sp.	4	2.5	1,198	0.5	green
13 Unidentified flagellate	4	2.5	74	0.0	
14 Aphanizomenon flos-aquae	3	2.0	2,786	1.2	bluegreen
15 Epithemia turgida	2	1.5	9,399	4.1	diatom
16 Cymbella minuta	2	1.5	818	0.4	diatom
17 Scenedesmus acuminatus	2	1.5	796	0.4	green
18 Cymbella tumida	2	1.5	5,529	2.4	diatom
19 Oocystis pusilla	1	1.0	318	0.1	green
20 Fragilaria crotonensis	1	1.0	17,338	7.6	diatom
21 Botryococcus braunii	1	1.0	8,492	3.7	green
22 Synedra radians	1	0.5	265	0.1	diatom
23 Tabellaria fenestrata	1	0.5	1,769	0.8	diatom
24 Gomphonema gracile	1	0.5	181	0.1	diatom
25 Ulothrix sp.	1	0.5	1,179	0.5	green
26 Synedra ulna	1	0.5	1,467	0.6	diatom
27 Fragilaria vaucheria	1	0.5	212	0.1	diatom
28 Asterionella formosa	1	0.5	1,297	0.6	diatom
29 Cosmarium sp.	1	0.5	155	0.1	green
30 Cymbella affinis	1	0.5	1,327	0.6	diatom
31 Staurastrum dejectum	1	0.5	295	0.1	green
32 Glenodinium sp.	1	0.5	516	0.2	dinoflagellate
Anabaena planctonica cells/mL =		295			
Anabaena planctonica heterocysts/mL =		9			
Aphanizomenon flos-aquae cells/mL =		44			
Microcystis aeruginosa cells/mL =		6,605			
<b>Aquatic Analysts</b>			<b>Sample ID:</b> KR76		

Phytoplankton Sample Analysis					
<b>Sample:</b> Tenmile Lake					
<b>Sample Station:</b> N16					
<b>Sample Depth:</b>					
<b>Sample Date:</b> 12-Sep-07					
<b>Total Density (#/mL):</b> 166					
<b>Total Biovolume (um<sup>3</sup>/mL):</b> 630,809					
<b>Trophic State Index:</b> 46.5					
<b>Species</b>	<b>Density #/mL</b>	<b>Density Percent</b>	<b>Biovolume um<sup>3</sup>/mL</b>	<b>Biovolume Percent</b>	<b>Group</b>
1 Anabaena planctonica	97	58.1	512,988	81.3	bluegreen
2 Aphanizomenon flos-aquae	25	14.7	30,932	4.9	bluegreen
3 Melosira granulata	12	7.4	35,105	5.6	diatom
4 Melosira ambigua	11	6.5	22,774	3.6	diatom
5 Melosira granulata angustissima	7	4.1	7,422	1.2	diatom
6 Microcystis aeruginosa	4	2.3	552	0.1	bluegreen
7 Rhodomonas minuta	3	1.8	61	0.0	cryptophyte
8 Ulothrix sp.	2	1.4	3,130	0.5	green
9 Asterionella formosa	2	0.9	1,013	0.2	diatom
10 Fragilaria crotonensis	2	0.9	12,888	2.0	diatom
11 Unidentified flagellate	1	0.5	15	0.0	
12 Dinobryon sertularia	1	0.5	91	0.0	dinoflagellate
13 Sphaerocystis schroeteri	1	0.5	3,437	0.5	green
14 Cryptomonas erosa	1	0.5	399	0.1	cryptophyte
Anabaena planctonica cells/mL =	2,803				
Anabaena planctonica heterocysts/mL =	15				
Aphanizomenon flos-aquae cells/mL =	491				
Microcystis aeruginosa cells/mL =	69				
<b>Aquatic Analysts</b>			<b>Sample ID:</b> KR77		



## APPENDIX II

Oregon Health Division  
Drinking water treatment guidance  
August 31, 2001

Contact Person: Ken Kauffman  
503-731-4015  
kenneth.w.kauffman@state.or.us

1. Treatment systems should consist of sand filtration followed by chlorination, followed by activated charcoal filtration. It is essential that sand filtration be done before disinfection to remove as many algal cells as possible without killing or rupturing them.
2. Chlorination systems should be capable of maintaining at least 1 ppm of chlorine residual for at least 20 minutes contact time before the water enters the activated charcoal system.
3. The final step in the process should be effective activated charcoal treatment to remove toxin remaining after the sand filtration and disinfection processes.
4. All treatment equipment used should meet NSF standard 53, and should be adequately sized to treat the maximum amount of water that you use. Treatment equipment needs regular monitoring and servicing to assure that it functions properly.
5. Ideally all water entering your home should be treated as recommended. It is possible to treat only water used in the kitchen, but this increases chances that animals or pets would inadvertently drink untreated water.

As more monitoring is done and toxin levels are measured this advisory may be altered. The advisory is to remain in effect until specifically changed or lifted by county and state health officials.

## FACT SHEET

# TOXIC *MICROCYSTIS* BLOOMS IN TENMILE LAKES

(information modified from Oregon Health Division Document: Hazards from *Microcystis aeruginosa* in Fresh Water – <http://www.ohd.hr.state.or.us/esc/docs/mafact.htm>)

### ➤ **What is a toxic bloom of *Microcystis aeruginosa*?**

*Microcystis aeruginosa* is a species of blue-green algae that grows naturally in many surface waters. In most bodies of fresh water and most weather conditions it does not pose a hazard to wildlife or human beings. However, under certain conditions (such as when the water is warm with abundant nutrients) *Microcystis aeruginosa* can grow more rapidly than normal. The result can be excessive numbers of large colonies that form floating masses on the water surface or that are dispersed within the water column. These occurrences are called "algal blooms". *Microcystis aeruginosa* can produce natural toxins (called microcystins) that are very potent, and these toxins are higher in concentration during bloom conditions. The microcystin toxins are produced and contained inside the *Microcystis* cells, and are released to the water when the cells die and disintegrate. Also, since the cells are very small, they can be ingested along with the water. Toxin levels in a water body tend to be higher near shorelines and at the surface of the water where animal and human contact is most likely.

### ➤ **What are the primary toxic effects of these blooms?**

The primary toxic effect of microcystins is on the liver. At very high doses, death of liver cells and destruction of blood vessels in the liver can result in serious injury and possibly death. Though less is known about the long-term effects of microcystin toxins, animal studies have shown these toxins can cause chronic liver damage and may promote the formation of liver tumors. These effects are more likely to occur if exposure is frequent over a long period of time.

The levels of toxin necessary to produce immediate or acute illness in humans and animals are much higher than levels that may cause chronic liver injury. Drinking water standards are usually based on chronic effects. Currently, there is no drinking water standard in the U.S. for microcystins. Canada, Australia, and Great Britain have developed a guideline level of 1 microgram toxin per liter of water, or 1 part per billion (1 ppb). During algal blooms, toxin levels can greatly exceed 1 ppb.

➤ **How is it determined when the water becomes safe once a bloom is reported?**

Changes in weather or in other conditions in a water body influence the growth of blue-green algae. Generally, cooler weather, rainfall, and reduced sunshine will lead to reductions in algal growth and toxin levels. Algal blooms generally peak and die off rapidly and toxin levels in the water decline over days or weeks. Only blue-green algae experts can distinguish visually between different kinds of algal growth, and are able to determine when blooms have disappeared. Testing of the water is the only way to be certain that toxin levels are no longer dangerous.

➤ **When does the Oregon Health Division Issue Warnings?**

**Drinking Water** -- When measured or estimated toxin levels reach 1 ug/l the Department of Human Services, Office of Public Health Systems issues public advisories or warnings. These will include warnings regarding the use of water for drinking or food preparation unless the water has been treated following specific guidelines for destroying and removing toxins. Animals should be kept away from water during periods when microcystin toxin levels exceed 1 ug/l, because drinking the water can cause serious or even fatal illness.

**Contact Recreation** -- If levels are high enough to pose hazards for swimming, water-skiing or other direct skin contact activities, the advisories will warn against water contact. Generally skin hazards occur where the water has a green or blue-green color or where there are visible clumps or mats of algae present in the water. When measured toxin levels reach 5 ug/L or cell counts reach 15,000 cells/ml, contact recreation is considered unsafe.

➤ **Can testing ensure that all areas of the lake are safe?**

No, due to the patchy nature of blue-green algal blooms it is possible for higher *Microcystis* densities (and therefore higher microcystin toxin concentrations) to be present in areas not sampled in a given survey, particularly along shorelines or during calm conditions of little to no wind. **Therefore, when a lake has a demonstrated history of algal toxicity or the presence of known toxin producing algal species, those utilizing the lake for drinking water should always follow Oregon Health Division recommendations for purification. In addition, recreational users should always avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance.**

➤ **Are domestic animals at risk during blooms?**

Yes, pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.

➤ **Is it safe to eat fish and other aquatic life?**

Clams, mussels, snails and other shellfish should not be eaten during microcystin advisory periods, but it is believed that fish can be safely eaten if they are cleaned and all internal organs discarded. Internal organs of such fish may be toxic even to animals.

➤ **How much does testing cost?**

Samples must be shipped to qualified laboratories for analysis. A microscopic determination to quantify the number of Microcystis colonies and cells costs \$90 per sample. A specialized test to analyze for the microcystin toxin concentration costs \$100 per sample (overnight shipping costs not included), and for anatoxins the cost is \$250/sample .

- NOTE: A fact sheet about microcystin toxin and its effects may be found on the Web at [www.dhs.state.or.us/publichealth/esc/docs/mufact.cfm](http://www.dhs.state.or.us/publichealth/esc/docs/mufact.cfm)