AQUATIC ECOSYSTEM SCIENCES, LLC

Jacob Kann, Ph.D.
Aquatic Ecologist

295 East Main St., Suite 7
Ashland, OR 97520
Voice: 541-482-1575
Fax: 541-552-1024
Email: jacobkann@aol.com

FAX

To:	Mike Mader/Jason – TLBP	From:	Jake Kann
Fax:	(503) 731-4077 541-759-3711	Pages:	13
Phone:		Date:	07-05-06
Re:	*Tenmile Sampling*	CC:	Dave Stone – OHD

Mike and Jason,

June 26th toxic algal cell count results for Tenmile Lakes are as follows:

STATION	DATE	*Microcystis* Cells (no./ml)	*Anabaena* Cells (no.ml)
S3	6-26-06	154	0
S8	6-26-06	0	26
N11	6-26-06	0	143
N16	6-26-06	99	1849

On this date only station N16 (1849 cells/ml) exceeded the WHO Alert Level 1 guideline (increased vigilance level for drinking water systems) of 500 cells ml^{-1} for *Anabaena flos-aquae*. *Microcystis aeruginosa* cells were present at very low levels at stations S3 and N16. Anabaena *was* present at low levels at S8 and S11. Counts for potentially toxigenic species were below the Alert Level 2 guideline of 2000 cells/ml at all stations (at which time OHS would issue a public alert for drinking water lakes and reservoirs).

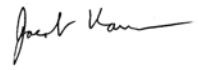
Stations S3, S8, and N11 were dominated by the diatoms Melosira sp., and *Fragilaria crotonensis.* Maximum *Microcystis aeruginosa* accounted for 1.7% of the biovolume at S3. Maximum *Anabaena* accounted for 19.1% of the biomass at N16. The cyanobacteria *Aphanizomenon flos-aquae* dominated (57.7%) the biovolume at N16. Various Chrysophytes, Cryptophytes, and Diatoms comprised the remainder of the biovolume at all stations.

Due to the patchy nature of blue-green algal blooms it is possible for higher *Microcystis aeruginosa* and *Anabaena flos-aquae* densities (and therefore higher microcystin toxin and anatoxin concentrations to be present in areas not sampled in this survey, particularly along shorelines or during calm conditions of little to no wind. Given the lakes' demonstrated history of toxic blooms, and the fact that all areas of the lake cannot be

tested at all times, those utilizing the lake for drinking water should always follow Oregon Health Division recommendations for purification (attached). In addition, recreational users should always avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance. Moreover, because pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.

Please call if you have any questions.

Sincerely,



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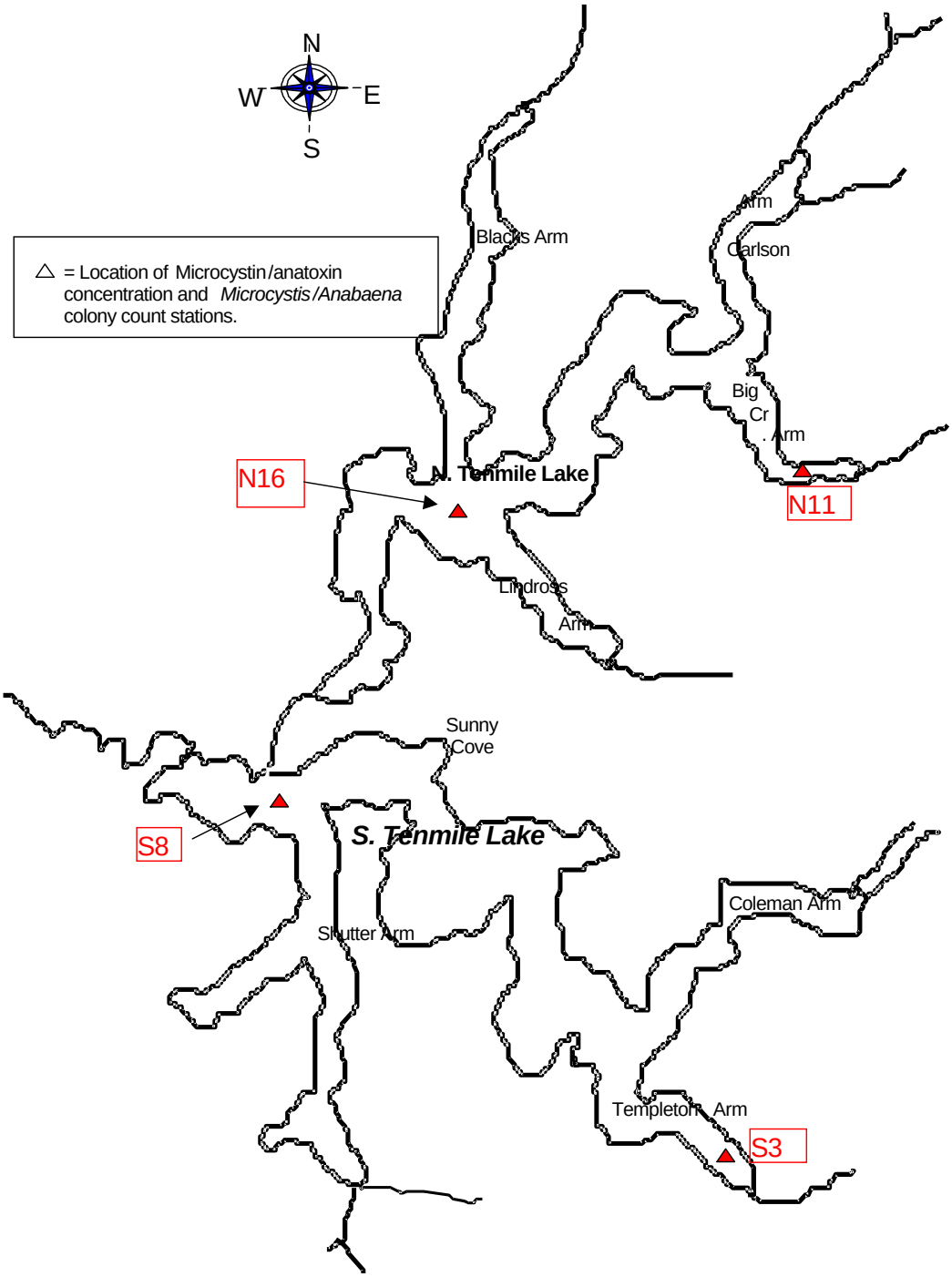
References for Alert Levels

Yoo, S.R., W.W. Carmichael, R.C. Hoehn, and S.E. Hrudy. 1995. Cyanobacterial (blue-green algal) toxins: a resource guide. AWWA Research Foundation and American Water Works Association. Denver, CO. 229 p. (ISBN 0-89867-824-2)

Falconer et al. 1999. Safe levels and safe practices. Pages 155-177 *in*: I. Chorus and J. Bartram, editors. *Toxic Cyanobacteria in water: a guide to their public health consequences*. World Health Organization Report. E & FN Spon, London and New York.

Year 2006 Tenmile Lakes Sample Site Locations



Phytoplankton Sample Analysis

Sample: Tenmile Lake
Sample Station: S3
Sample Depth:
Sample Date: 26-Jun-06

Total Density (#/mL): 33
Total Biovolume (um^3/mL): 74,472
Trophic State Index: 31.2

	Species	Density #/mL	Density Percent	Biovolume um^3/mL	Biovolume Percent
1	Melosira granulata	7	22.0	29,509	39.6
2	Microcystis aeruginosa	7	20.0	1,229	1.7
3	Aphanizomenon flos-aquae	5	16.0	3,704	5.0
4	Melosira granulata angustissima	3	10.0	5,679	7.6
5	Gloeotrichia echinulata	2	6.0	17,310	23.2
6	Rhodomonas minuta	1	4.0	27	0.0
7	Dinobryon sertularia	1	4.0	159	0.2
8	Glenodinium sp.	1	2.0	468	0.6
9	Eudorina elegans	1	2.0	4,169	5.6
10	Synedra rumpens	1	2.0	94	0.1
11	Fragilaria crotonensis	1	2.0	8,980	12.1
12	Navicula capitata	1	2.0	321	0.4
13	Sphaerocystis schroeteri	1	2.0	374	0.5
14	Nitzschia sp.	1	2.0	80	0.1
15	Ankistrodesmus falcatus	1	2.0	17	0.0
16	Asterionella formosa	1	2.0	2,352	3.2

Microcystis aeruginosa cells/mL = 154

Aphanizomenon flos-aquae cells/mL = 64
Aphanizomenon flos-aquae heterocysts/mL = 1

Gloeotrichia echinulata cells/mL = 255
Gloeotrichia echinulata heterocysts/mL = 7

Aquatic Analysts **Sample ID:** JU50

Phytoplankton Sample Analysis

Sample: Tenmile Lake
Sample Station: S8
Sample Depth:
Sample Date: 26-Jun-06

Total Density (#/mL): 309
Total Biovolume (um^3/mL): 603,402
Trophic State Index: 46.2

	Species	Density #/mL	Density Percent	Biovolume um^3/mL	Biovolume Percent
1	Melosira granulata angustissima	158	51.3	277,027	45.9
2	Aphanizomenon flos-aquae	71	23.1	94,245	15.6
3	Melosira granulata	47	15.4	180,226	29.9
4	Melosira ambigua	11	3.4	37,296	6.2
5	Dinobryon sertularia	8	2.6	942	0.2
6	Asterionella formosa	5	1.7	11,609	1.9
7	Ankistrodesmus falcatus	3	0.9	66	0.0
8	Ochromonas sp.	3	0.9	224	0.0
9	Anabaena flos-aquae	3	0.9	1,768	0.3

Aphanizomenon flos-aquae cells/mL = 1,496
Aphanizomenon flos-aquae heterocysts/mL = 29

Anabaena flos-aquae cells/mL = 26
Anabaena flos-aquae heterocysts/mL = 3

Aquatic Analysts

Sample ID: JU51

Phytoplankton Sample Analysis

Sample:	Tenmile Lake
Sample Station:	N11
Sample Depth:	
Sample Date:	26-Jun-06

Total Density (#/mL):	2,467
Total Biovolume (um^3/mL):	232,852
Trophic State Index:	39.4

	Species	Density #/mL	Density Percent	Biovolume um^3/mL	Biovolume Percent
1	Unidentified flagellate	2,404	97.4	48,087	20.7
2	Dinobryon sertularia	19	0.8	2,253	1.0
3	Anabaena flos-aquae	8	0.3	9,584	4.1
4	Fragilaria capucina mesolepta	4	0.2	53,640	23.0
5	Asterionella formosa	4	0.2	12,958	5.6
6	Aphanizomenon flos-aquae	4	0.2	5,301	2.3
7	Synedra ulna	2	0.1	4,186	1.8
8	Gomphonema subclavatum	2	0.1	2,524	1.1
9	Oocystis pusilla	2	0.1	454	0.2
10	Cocconeis placentula	2	0.1	968	0.4
11	Melosira ambigua	2	0.1	9,912	4.3
12	Gloeotrichia echinulata	2	0.1	28,608	12.3
13	Fragilaria crotonensis	2	0.1	53,009	22.8
14	Mallomonas sp.	2	0.1	799	0.3
15	Chaetoceros sp.	2	0.1	200	0.1
16	Ankistrodesmus falcatus	2	0.1	53	0.0
17	Achnanthes minutissima	2	0.1	316	0.1

Gloeotrichia echinulata cells/mL =	421
Gloeotrichia echinulata heterocysts/mL =	105
Aphanizomenon flos-aquae cells/mL =	84
Aphanizomenon flos-aquae heterocysts/mL =	1
Anabaena flos-aquae cells/mL =	143
Anabaena flos-aquae heterocysts/mL =	11

Aquatic Analysts **Sample ID:** JU52

Phytoplankton Sample Analysis

Sample: Tenmile Lake
Sample Station: N16
Sample Depth:
Sample Date: 26-Jun-06

Total Density (#/mL): 445
Total Biovolume (um^3/mL): 649,295
Trophic State Index: 46.7

	Species	Density #/mL	Density Percent	Biovolume um^3/mL	Biovolume Percent
1	Aphanizomenon flos-aquae	313	70.4	374,894	57.7
2	Anabaena flos-aquae	56	12.6	123,916	19.1
3	Melosira granulata angustissima	46	10.4	85,387	13.2
4	Melosira granulata	10	2.2	16,319	2.5
5	Melosira ambigua	7	1.5	31,069	4.8
6	Microcystis aeruginosa	3	0.7	791	0.1
7	Ochromonas sp.	3	0.7	280	0.0
8	Dinobryon sertularia	3	0.7	392	0.1
9	Oocystis lacustris	3	0.7	16,247	2.5

Aphanizomenon flos-aquae cells/mL = 5,951
Aphanizomenon flos-aquae heterocysts/mL = 73

Microcystis aeruginosa cells/mL = 99

Anabaena flos-aquae cells/mL = 1,849
Anabaena flos-aquae heterocysts/mL = 92
Anabaena flos-aquae akinetes/mL = 3

Aquatic Analysts **Sample ID:** JU53

July 31, 2006

Oregon Health Division **Contact Person: Ken Kauffman**
Drinking water treatment guidance **503-731-4015**
August 31, 2001 **kenneth.w.kauffman@state.or.us**

1. Treatment systems should consist of sand filtration followed by chlorination, followed by activated charcoal filtration. It is essential that sand filtration be done before disinfection to remove as many algal cells as possible without killing or rupturing them.

2. Chlorination systems should be capable of maintaining at least 1 ppm of chlorine residual for at least 20 minutes contact time before the water enters the activated charcoal system.

3. The final step in the process should be effective activated charcoal treatment to remove toxin remaining after the sand filtration and disinfection processes.

4. All treatment equipment used should meet NSF standard 53, and should be adequately sized to treat the maximum amount of water that you use. Treatment equipment needs regular monitoring and servicing to assure that it functions properly.

5. Ideally all water entering your home should be treated as recommended. It is possible to treat only water used in the kitchen, but this increases chances that animals or pets would inadvertently drink untreated water.

As more monitoring is done and toxin levels are measured this advisory may be altered. The advisory is to remain in effect until specifically changed or lifted by county and state health officials.

FACT SHEET

TOXIC *MICROCYSTIS* BLOOMS IN TENMILE LAKES

(information modified from Oregon Health Division Document: Hazards from *Microcystis aeruginosa* in Fresh Water – http://www.ohd.hr.state.or.us/esc/docs/mafact.htm)

➢ What is a toxic bloom of *Microcystis aeruginosa*?

Microcystis aeruginosa is a species of blue-green algae that grows naturally in many surface waters. In most bodies of fresh water and most weather conditions it does not pose a hazard to wildlife or human beings. However, under certain conditions (such as when the water is warm with abundant nutrients) *Microcystis aeruginosa* can grow more rapidly than normal. The result can be excessive numbers of large colonies that form floating masses on the water surface or that are dispersed within the water column. These occurrences are called "algal blooms". *Microcystis aeruginosa* can produce natural toxins (called microcystins) that are very potent, and these toxins are higher in concentration during bloom conditions. The microcystin toxins are produced and contained inside the *Microcystis* cells, and are released to the water when the cells die and disintegrate. Also, since the cells are very small, they can be ingested along with the water. Toxin levels in a water body tend to be higher near shorelines and at the surface of the water where animal and human contact is most likely.

➢ What are the primary toxic effects of these blooms?

The primary toxic effect of microcystins is on the liver. At very high doses, death of liver cells and destruction of blood vessels in the liver can result in serious injury and possibly death. Though less is known about the long-term effects of microcystin toxins, animal studies have shown these toxins can cause chronic liver damage and may promote the formation of liver tumors. These effects are more likely to occur if exposure is frequent over a long period of time.

The levels of toxin necessary to produce immediate or acute illness in humans and animals are much higher than levels that may cause chronic liver injury. Drinking water standards are usually based on chronic effects. Currently, there is no drinking water standard in the U.S. for microcystins. Canada, Australia, and Great Britain have developed a guideline level of 1 microgram toxin per liter of water, or 1 part per billion (1 ppb). During algal blooms, toxin levels can greatly exceed 1 ppb.

➢ How is it determined when the water becomes safe once a bloom is reported?

Changes in weather or in other conditions in a water body influence the growth of blue-green algae. Generally, cooler weather, rainfall, and reduced sunshine will lead to reductions in algal growth and toxin levels. Algal blooms generally peak and die off rapidly and toxin levels in the water decline over days or weeks. Only blue-green algae experts can distinguish visually between different kinds of algal growth, and are able to determine when blooms have disappeared. Testing of the water is the only way to be certain that toxin levels are no longer dangerous.

➢ When does the Oregon Health Division Issue Warnings?

Drinking Water -- When measured or estimated toxin levels reach 1 ug/l the Department of Human Services, Office of Public Health Systems issues public advisories or warnings. These will include warnings regarding the use of water for drinking or food preparation unless the water has been treated following specific guidelines for destroying and removing toxins. Animals should be kept away from water during periods when microcystin toxin levels exceed 1 ug/l, because drinking the water can cause serious or even fatal illness.

Contact Recreation -- If levels are high enough to pose hazards for swimming, water-skiing or other direct skin contact activities, the advisories will warn against water contact. Generally skin hazards occur where the water has a green or blue-green color or where there are visible clumps or mats of algae present in the water. When measured toxin levels reach 5 ug/L or cell counts reach 15,000 cells/ml, contact recreation is considered unsafe.

➢ Can testing ensure that all areas of the lake are safe?

No, due to the patchy nature of blue-green algal blooms it is possible for higher *Microcystis* densities (and therefore higher microcystin toxin concentrations) to be present in areas not sampled in a given survey, particularly along shorelines or during calm conditions of little to no wind. **Therefore, when a lake has a demonstrated history of algal toxicity or the presence of known toxin producing algal species, those utilizing the lake for drinking water should always follow Oregon Health Division recommendations for purification. In addition, recreational users should always avoid contact with water whenever noticeable surface concentrations of algae are evident or when the lake has an obvious green to blue-green appearance.**

➢ Are domestic animals at risk during blooms?

Yes, pets or other domestic animals are the most likely to ingest contaminated water, these animals should not be allowed access to the lakeshore whenever either noticeable surface concentrations of algae or an obvious green to blue-green appearance is evident.

➢ Is it safe to eat fish and other aquatic life?

Clams, mussels, snails and other shellfish should not be eaten during microcystin advisory periods, but it is believed that fish can be safely eaten if they are cleaned and all internal organs discarded. Internal organs of such fish may be toxic even to animals.

➢ How much does testing cost?

Samples must be shipped to qualified laboratories for analysis. A microscopic determination to quantify the number of Microcystis colonies and cells costs $90 per sample. A specialized test to analyze for the microcystin toxin concentration costs $100 per sample (overnight shipping costs not included), and for anatoxins the cost is $250/sample .

- NOTE: A fact sheet about microcystin toxin and its effects may be found on the Web at www.dhs.state.or.us/publichealth/esc/docs/mafact.cfm